

Paul Pfeiffer^{1,2}, Federico José Barreda Tomás^{2,3}, Jiameng Wu^{1,2}, Jan-Hendrik Schleimer^{1,2}, Imre Vida^{2,3}, Susanne Schreiber^{1,2}

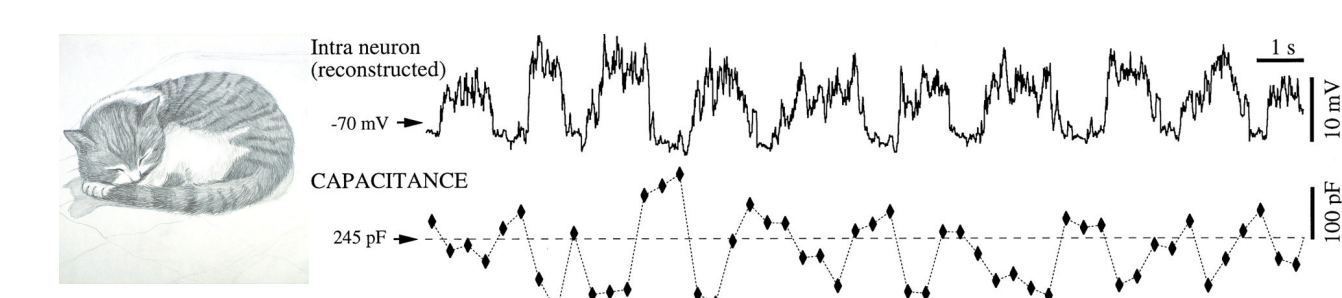
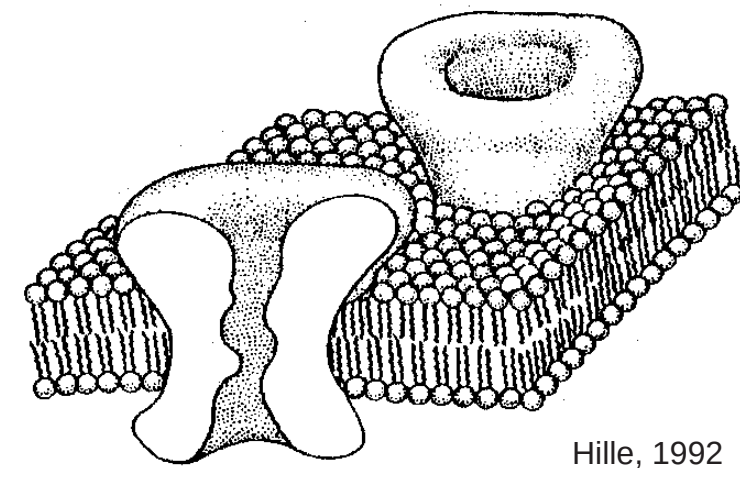
¹ Institute for Theoretical Biology, Humboldt-Universität zu Berlin, Philippstr. 13, Haus 4, 10115 Berlin, Germany

² Bernstein Center for Computational Neuroscience, Humboldt-Universität zu Berlin, Philippstr. 13, Haus 6, 10115 Berlin, Germany

³ Institute for Integrative Neuroanatomy, Charité-Universitätsmedizin Berlin, Charitéplatz 1, 10117 Berlin, Germany

Motivation

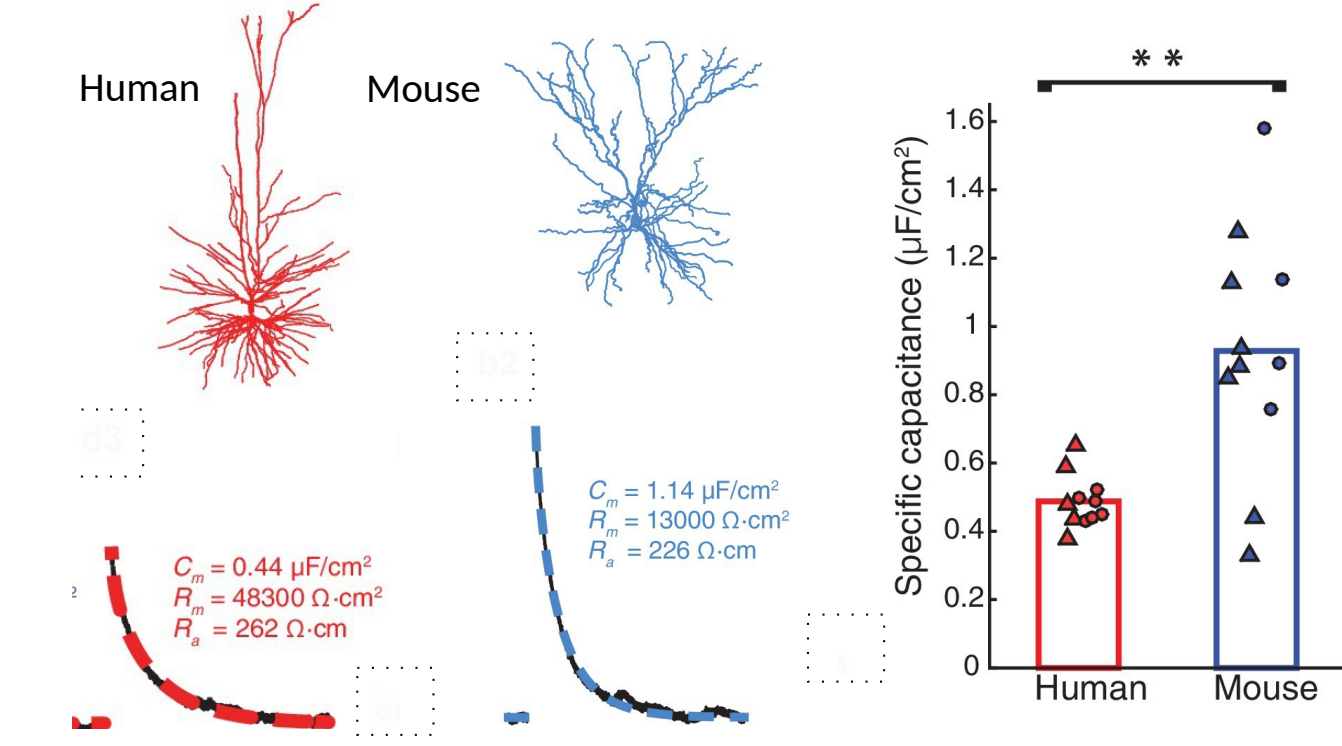
Membrane capacitance translates ionic and synaptic membrane currents into voltage responses. Importantly, it sets the membrane time constant and hence fundamentally affects neural signal transmission and processing.



C_m is not a (boring) constant

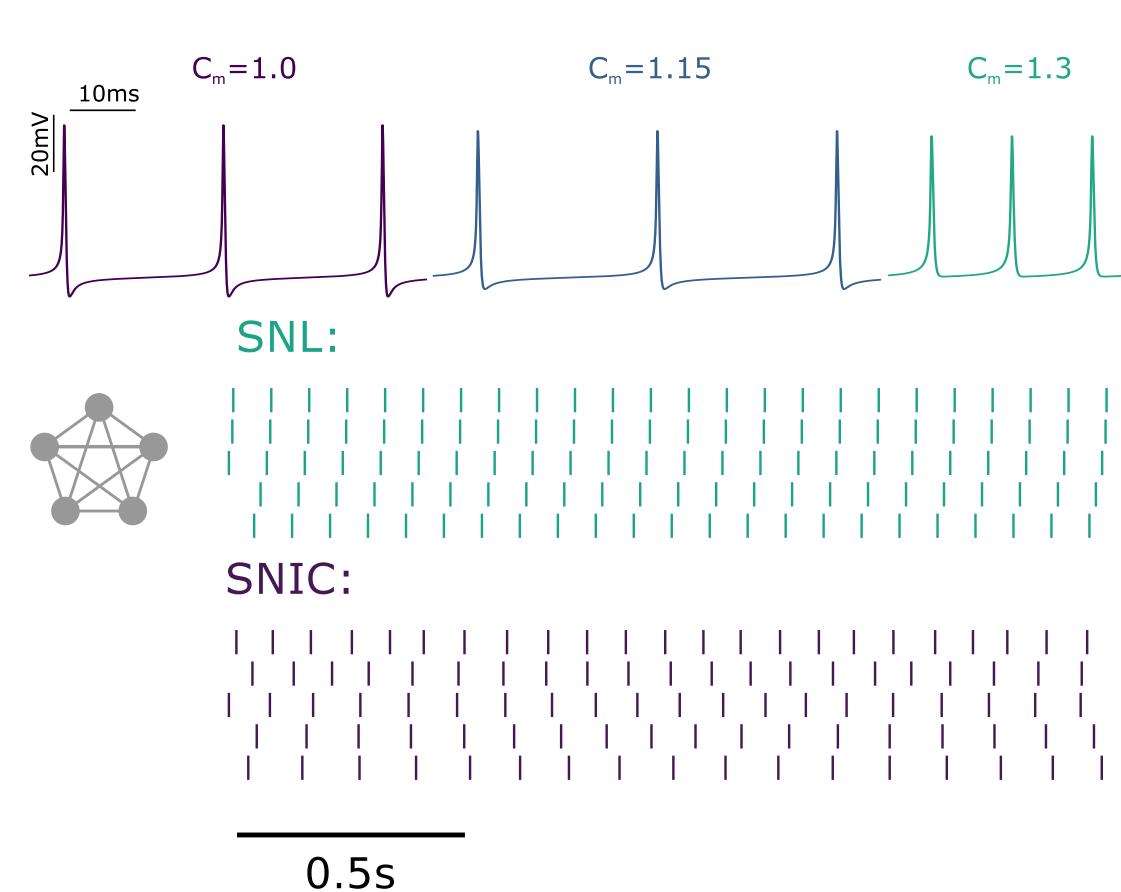
The membrane capacitance of neurons and glia changes during physiological states like slow wave sleep probably by swelling¹. An altered membrane capacitance is also observed in pathological conditions like Alzheimers².

The specific membrane capacitance is commonly assumed to be a universal constant of $1 \mu\text{F}/\text{cm}^2$. However, it seems to vary across species and can change during development³.



Capacitance as a critical switch

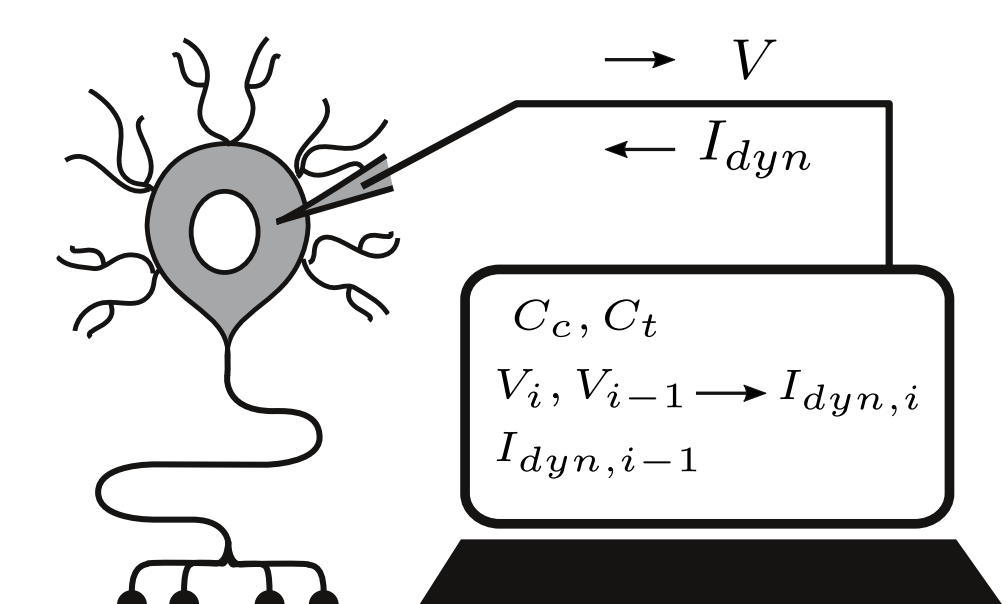
Changes in capacitance alter the relative speed of ionic currents and the membrane potential. Modelling predicts that capacitance changes, like temperature increase or exhaustion of ionic concentration gradients, can induce a switch in action potential generation, which affects the coding ability of single neurons, but also network phenomena like synchronization⁴.



Control of capacitance in experiments

Manipulation of capacitance by pharmacological or genetical means is hard to control. Here, we propose the capacitance clamp, which uses the dynamic clamp to precisely and robustly control the capacitance of an excitable cell.

Clamp capacitance via dynamic clamp

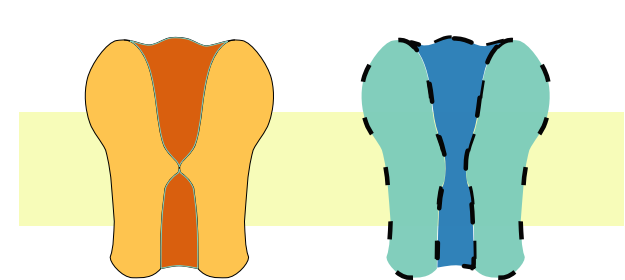


Dynamic clamp

Fast feedback loop between recorded voltage and injected current to mimic changes in the electrical properties of a neuron⁵.

$$C_m \frac{dV}{dt} = I(V) + I_{dyn}$$

Classical conductance clamp



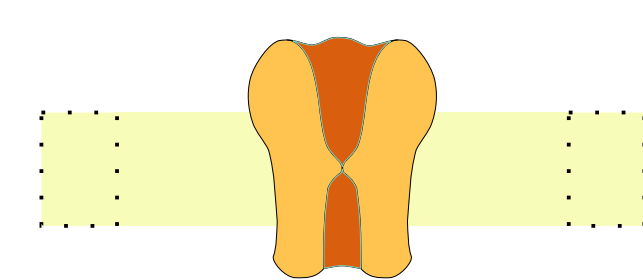
Aim: mimic additional ion channels

$$C_m \frac{dV}{dt} = I(V) + g(V - E)$$

$$\Rightarrow I_{dyn} = g(V - E)$$

$$\approx g(V_i - E)$$

Proposal capacitance clamp



Aim: mimic a modified membrane

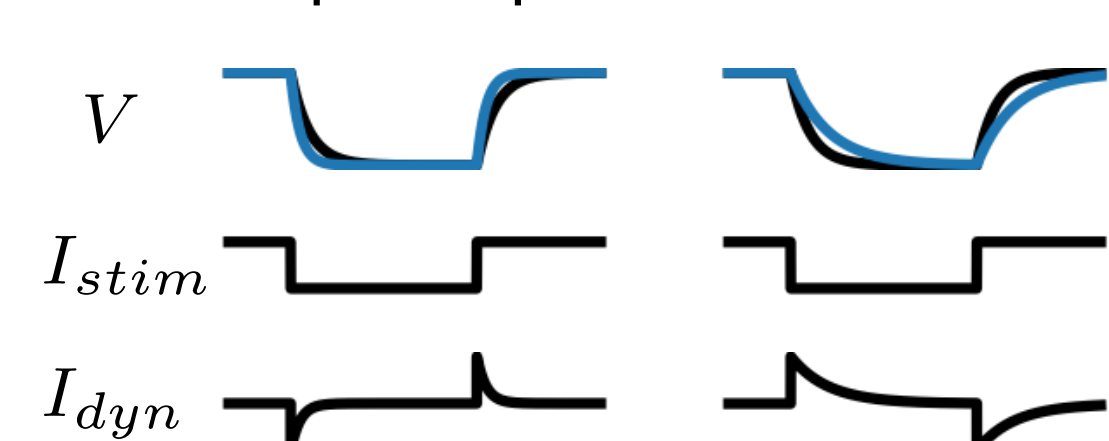
$$C_t \frac{dV}{dt} = I(V)$$

$$\Rightarrow I_{dyn} = \frac{C_c - C_t}{C_t} I(V)$$

$$\approx \frac{C_c - C_t}{C_t} \left(\frac{V_i - V_{i-1}}{C_c \Delta t} - I_{dyn, i-1} \right)$$

$C_t < C_c$
speed up

$C_t > C_c$
slow down



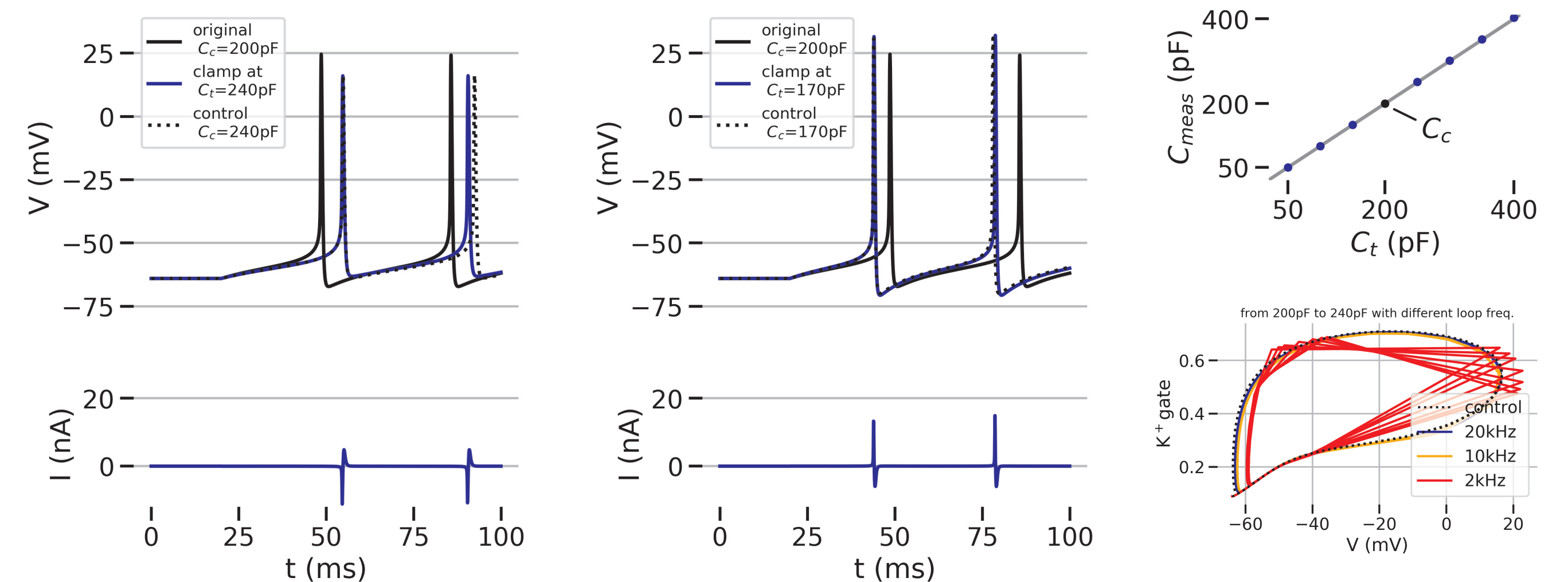
Capacitance clamp allows the experimenter to precisely control the capacitance of an excitable cell.

Only requires to measure the cell capacitance C_c ; in particular no needs for a detailed model of the present ionic currents.

Results

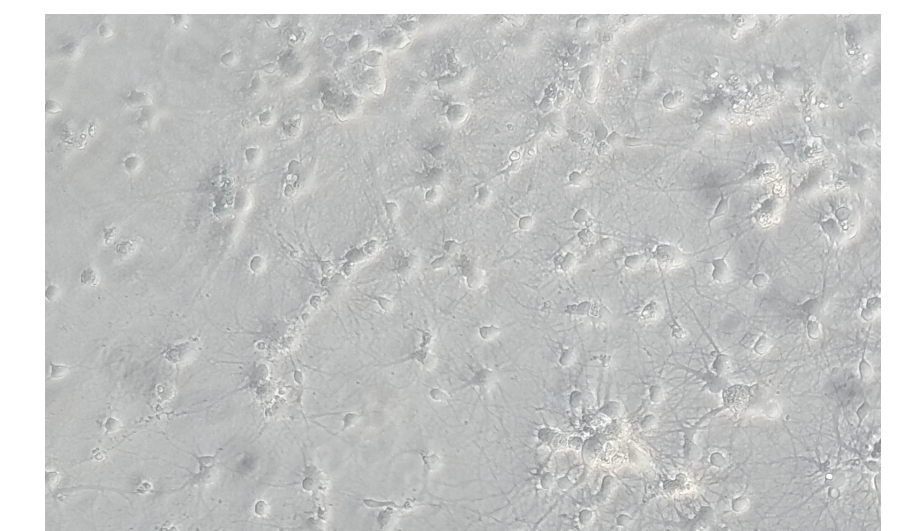
Capacitance clamp in silico

We simulated the capacitance clamp with conductance-based neuron models and compared the electrical activity of a neuron clamped at a target capacitance with a "control" neuron at this capacitance.

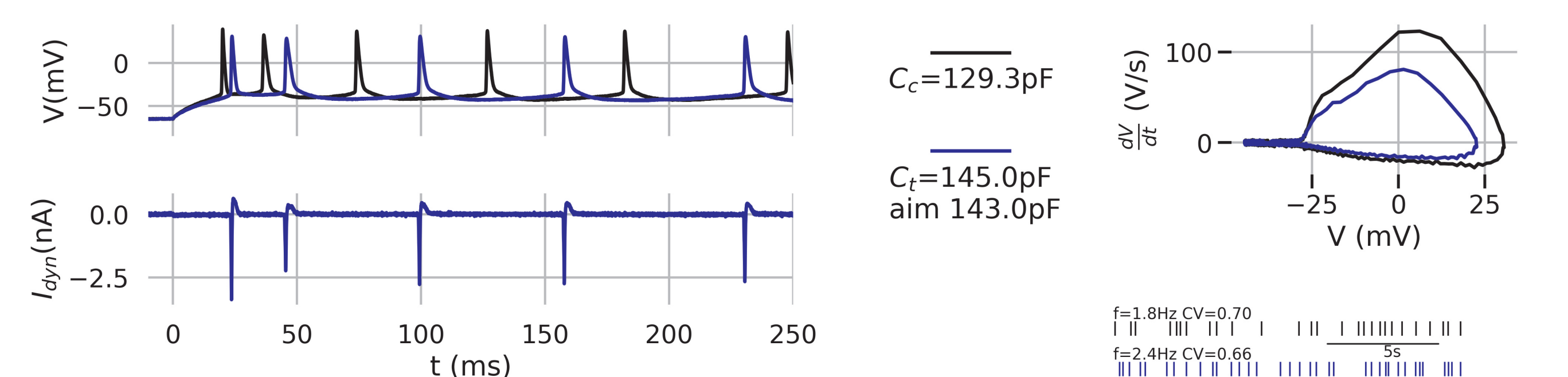


Capacitance clamp in vitro

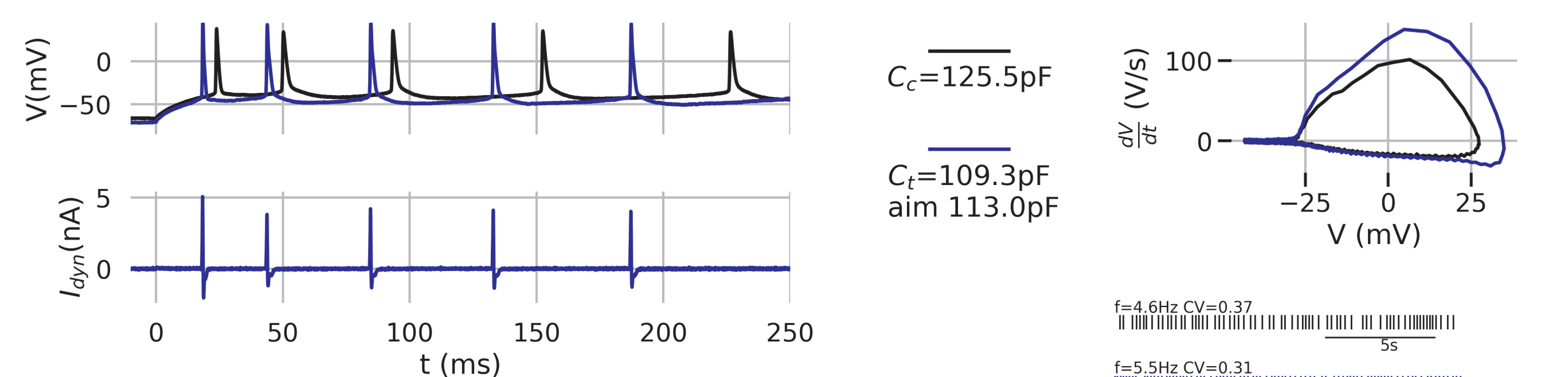
We tested the capacitance clamp in cultured hippocampal neurons. We used step pulses to measure the capacitance and then compared spiking close to the threshold at different capacitances. The software for dynamic clamp was RELACS⁶.



Increasing capacitance $C_t > C_c$



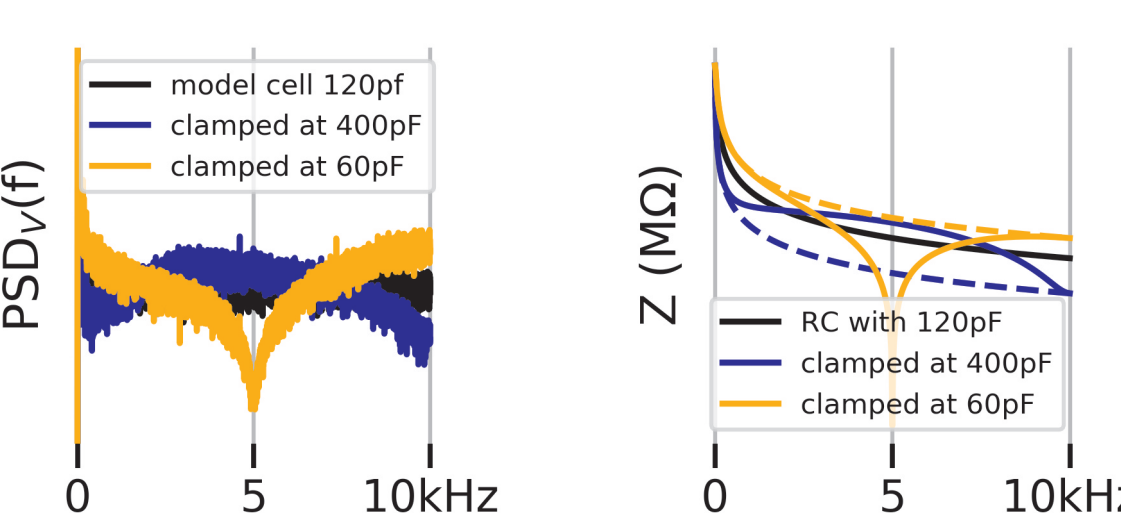
Decreasing capacitance $C_t < C_c$



Outlook

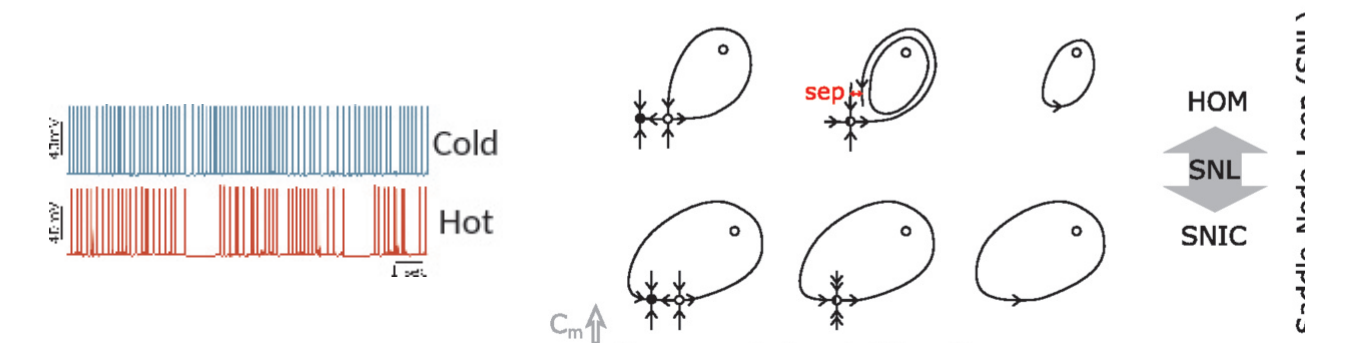
We present a novel tool for electrophysiology: the capacitance clamp provides reliable control over the capacitance of a neuron and thereby opens a new way to study the temporal dynamics of excitable cells.

Robustness



We work on an optimal filter design to make the capacitance clamp less noisy and more robust against imperfect electrode compensation.

Applications



We apply the capacitance clamp to study how close neurons are to a critical switch in spike generation, the SNL bifurcation, which has widespread implications from coding to epilepsy.

Contact

@: pfeiffpa@hu-berlin.de (Paul Pfeiffer) and s.schreiber@hu-berlin.de (Susanne Schreiber)
web: neuron-science.de

Acknowledgments

This work was supported by BMBF (01GQ0901, 01GQ1403) and DFG (GRK 1589/2). We are grateful to Jan Benda and his lab for their dedicated support with dynamic clamp. We thank Susana Contreras and Dr. Janina Hesse for valuable discussions and remarks.

References

- Amzica, F. & Neckelmann, D. Membrane Capacitance of Cortical Neurons and Glia During Sleep Oscillations and Spike-Wave Seizures.
- Brown, J. T. et al. Altered intrinsic excitability of hippocampal CA1 pyramidal neurons in aged PDAPP mice. *Front. Cell. Neurosci.* 9, 1–14 (2015).
- Eyal, G. et al. Unique membrane properties and enhanced signal processing in human neocortical neurons. *Elife* 5, 1–18 (2016).
- Hesse, J., Schleimer, J. H. & Schreiber, S. Qualitative changes in phase-response curve and synchronization at the saddle-node-loop bifurcation. *Phys. Rev. E* 95, (2017).
- Sharp, A. A., O'Neil, M. B., Abbott, L. F. & Marder, E. The dynamic clamp: artificial conductances in biological neurons. *Trends Neurosci.* 16, 389–394 (1993).
- www.relacs.net